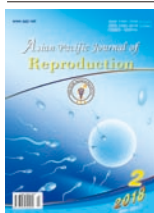


Asian Pacific Journal of Reproduction



2
2018
Mar. Vol. 7



Asian Pacific Journal of Reproduction

Journal homepage: www.apjr.net



doi: 10.4103/2305–0500.228013

©2018 by the Asian Pacific Journal of Reproduction. All rights reserved.

Germline cells derived from mesenchymal stem cells, with the focus on Wharton's jelly

Hossein Yazdekhesti¹, Jalil Hosseini², Zahra Rajabi^{3,4}, Maryam Hosseinzadeh Shirzeyli⁵, Fereshte Aliakbari⁵✉

¹Department of Cell Biology, Center for Research in Contraceptive and Reproductive Health, University of Virginia, Charlottesville, VA 22908, USA

²Infertility and Reproductive Health Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

³Department of Anatomy, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran

⁴Department of Biomedical Engineering, University of Virginia, Charlottesville, VA 22908, USA

⁵Department of Biology and Anatomical Sciences, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

ARTICLE INFO

Article history:

Received 15 November 2017

Revision 8 December 2017

Accepted 25 December 2017

Available online 20 March 2017

Keywords:

Mesenchymal stem cell

Bone marrow stem cell

Umbilical cord

Wharton's jelly

ABSTRACT

Previous attempts have indicated that mesenchymal stem cells (MSCs) are a valuable source and candidate and new approach for tissue engineering and reproductive medicine. MSCs have this potential to be induced and differentiated in an appropriate *in vivo* and *in vitro* condition toward various cell lineages and then they can be applied in cell therapies and clinical applications. During recent two decades, various sources have demonstrated they are a great source for MSCs, including bone marrow, the human umbilical cord as well as Wharton's jelly. Due to discarding after birth, easily accessible cells and less ethical concerns, these cells have attracted more and more scientists' attention. Infertility and reproduction diseases have provided special opportunity to examine the efficiency of MSCs in this kind of application. Based on recent investigations, MSCs embedded in Wharton's jelly tissue are more appealing for cell therapies, especially in infertility treatment purposes. So, differentiation of MSCs embedded in Wharton's jelly tissue into germ layer cells for cell-based therapy purposes is now under intensive study.

1. Introduction

Mesenchymal stem cells (MSCs) are a valuable source for clinical application of cells and tissue engineering[1–3]. Source of variations arise from whether the cells were derived from allogeneic or autologous sources. Autologous sources are more appreciated because they eliminate issues such as contamination and risk of malignancy[4]. The allogeneic application brings some

others problems such as possible ineffectiveness[5]. There is a body of reports showing that MSCs have a particular function in *in vitro* and *in vivo* condition. They display immunomodulatory functions and inhibit T-lymphocyte proliferation and activation and induced by cellular factors[6,7], and respond to injury or stress, just like the respond of immune system cells to pathogen exposure[8,9], participation in regeneration, immune cell activation

✉Corresponding author: Fereshte Aliakbari, Department of Biology and Anatomical Sciences, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

E-mail: fereshtehaliakbari@yahoo.com

Tel: 09138908329

Fax: 02122716383

First author: Hossein Yazdekhesti, Department of Cell Biology, Center for Research in Contraceptive and Reproductive Health, University of Virginia, Charlottesville, VA 22908, USA.

This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial-Share Alike 3.0 License, which allows others to remix, tweak and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

©2018 Asian Pacific Journal of Reproduction Produced by Wolters Kluwer- Medknow

How to cite this article: Hossein Yazdekhesti, Jalil Hosseini, Zahra Rajabi, Maryam Hosseinzadeh Shirzeyli, Fereshte Aliakbari. Germline cells derived from mesenchymal stem cells, with the focus on Wharton's jelly. *Asian Pac J Reprod* 2018; 7(2): 49-55.

or suppression, angiogenesis, remodeling, bactericidal activity and cellular recruitment[10]. MSCs also can be found in different adults and birth-associated tissues. This review study focused on some of the standard features of MSCs from various sources and their differentiation capacity toward germ line cells with an emphasis on Wharton's jelly.

2. Mesenchymal stromal cells

Mesenchymal stromal cells or stem cells are a great source of multipotent stem cells with self-renewal capacity, which have the capacity of differentiation into various cell lineages and be transdifferentiated toward astrocytes-like cells, hepatocytes and neural cells *in vitro* as well[11,12]. Due to their capacities, they are always utilized in tissue regenerative medicine and transplantation studies[13–17].

MSCs are derived from various tissues including bone marrow, adipose tissue, adult and fetal tissues and Wharton's jelly of the umbilical cord (Figure 1). They are undifferentiated cells that can be mostly found in embryonic and extraembryonic tissues[18]. The embryonic tissues containing MSCs are including spleen, fetal bone marrow, pancreas, lung, liver, and peripheral blood and the extra-embryonic structures such as umbilical cord, umbilical cord blood, amniotic fluid, placenta and amnion, are containing mesenchymal stem cells[2,19].

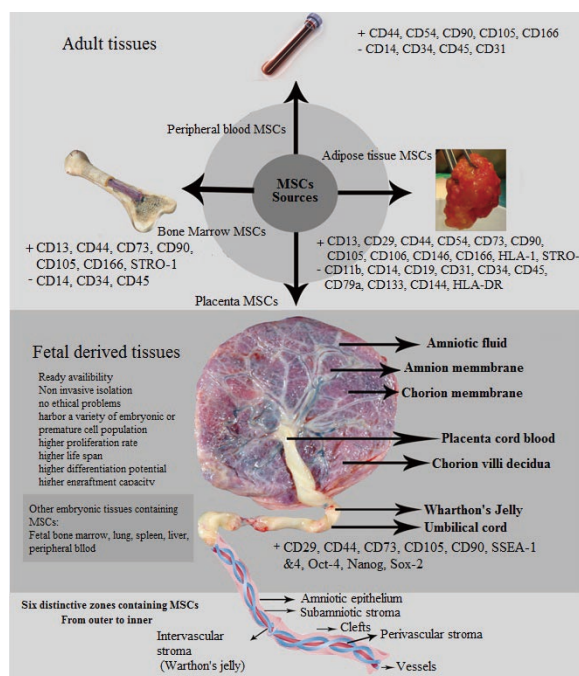


Figure 1. MSCs features from some sources and their differentiation capacity toward germline cells with an emphasis on Wharton's jelly.

Based on differentiation potential and proliferation capacity, there are variations between fetal and adult MSCs. Fetal MSCs have some priorities and advantages over other sources such as faster doubling time than adult MSCs, greater expansion capacity *in vitro* as well as longer telomeres[20]. Fetal MSCs don't have the properties of immune suppression, for instance, lack of class II human leukocyte antigens (HLA II), but they seem to synthesize HLA-G, in contrast to adult MSCs in which HLA II is present and HLA-G is absent[21]. Overall, fetal MSCs are secreting a slightly different cocktail of cytokine than adult MSCs (Table 1)[22].

Table 1

Markers of BM-MSC, umbilical cord, and Wharton's jelly.

Marker	BM-MSC	Umbilical cord	Wharton's jelly	References
HLA-A	+	+	+	[68–70]
HLA-B	+	+	+	[68,71–73]
HLA-C	+	+	+	[68,71,74,75]
HLA-DR	-	-	-	[69,70,74,76]
HLA-G	+	-	+	[54,77,78]
CD29	+	+	+	[11,27,59,79]
CD44	+	+	+	[11,59,78]
CD73	+	+	+	[11,59,73,78]
CD105	+	+	+	[11,59,78]
CD53	+	+	+	[11,59,78]
CD54 ICAM-1	+	-/+a	n.a	[79–82]
CD56	-	-	-	[11,59,70]
CD58	+	-	+	[11,59]
CD106 VCAM-1	+	-/+	+	[78,80,81,83]
CD166 ALCAM	+	+	+	[69,78,81]
SSEA-1	+	-	+	[59,78]
Oct-04	+	-/+	+	[59,70,78]
Nanog	+	+	+	[59,70,84]
Sox-2	+	+	+	[59,84]

Two methods have been applied for isolation of MSCs including enzymatic digestion and tissue culture (insert method). In order to perform enzymatic digestion, after the membrane and veins have been removed, collagenase and trypsin are routinely utilized to digest the umbilical cord tissue. It has been proposed that trypsin and collagenase might damage the Wharton's jelly, however, this method has increased the outcomes of obtained cells[23]. Meanwhile, enzymatic digestion is unaffordable with a high risk of contamination and takes more time to perform and is not easy to control[24]. Mechanical digestion of the cord is an essential step before the onset of enzymatic digestion[25,26]. The common point of digestion is the use of collagenase-containing caseinase, clostripain, and tryptic activities. Type I collagenase is routinely used for the isolation of stromal cells[27]. A combination of collagenase with hyaluronidase is critically important because it facilitates the outcomes of matrix digestion and shortens the time required for isolation process[26].

Different independent groups have reported their successful

isolated MSCs from umbilical cord using culture method[28,29]. Explant of tissue fragments is one of the most primitive techniques in cell isolation and propagation *in vitro*. This approach affects the quantity and quality of the isolated cells, but the tissue size should be small enough for freely gases and nutrients diffusion[30]. The primary explanted culture success rate is directly dependent on the migratory ability of the cell type[31].

3. MSCs derived from bone marrow

The first source which was claimed to comprise MSCs was bone marrow[32], and MSCs were obtained from bone marrow by Friedenstein's team for the first time[33]. They described these cells as a population of cells similar to fibroblast-like colonies with the capacity of differentiating toward multiple mesenchymal lineages and then Caplan *et al*[34] called these cells as "mesenchymal stem cells". Finally, Horwitz *et al*[35] recently referred these cells as "multipotent mesenchymal stromal cells".

The procedure of sampling from bone marrow is an annoying and invasive procedure[36], and along with aging and adolescence, the bone marrow-MSCs (BM-MSCs) number decreases[20]. Meanwhile, this should be always considered in mind that the risk of viral contamination during the isolation of MSCs from bone marrow is still present[37]. Due to all of these reasons, the application of bone marrow in cell therapy procedures as a great source of MSCs has been limited. Therefore, the applications of other sources which have MSCs with a higher proliferative and differentiation potency and lower risk of viral contamination have been considered.

BM-MSCs have the ability of self-renewal and differentiate into connective tissues cells such as adipocytes, osteoblasts as well as chondrocytes[11,38,39]. They express various cell surface markers including CD29, CD44, CD73, CD90, CD105, CD166, CD49e, CD51, CD54, CD59, CD71 and CD200, however there are some other surface markers which BM-MSCs do not express (such as CD14, CD31, CD34, CD45, CD79, CD86, CD117 and glycoporphin A) (Table 1)[11]. Characterizations of BM-MSCs and their non-tumorigenic properties have made them a suitable candidate for human therapeutic applications, particularly in degenerative diseases by autologous cell transplantation[40]. Also, they do not induce proliferation of T-lymphocyte *in vitro* and some reports have shown that they prevent the T-cell responses to mitogenic and antigenic stimuli. They don't have the capacity to stimulate B cells and are resistant to lysis which has been mediated by the natural killer cell[41,42]. Di Nicola *et al*[6] indicated that transforming growth factor- β and hepatocyte growth factor block T-cell expansion in mixed lymphocyte reaction and T-lymphocytes are suppressed by BM-MSCs, and they couldn't enter apoptosis.

Johnson *et al*[43] reported that there is some evidence showing that oocyte might generate from bone marrow in adult mammalian ovaries. Their results revealed that bone marrow is a considerable

origin of germ cells which lead to the continuation of oocyte production during adulthood. Moreover, Bukovsky *et al*[44,45] observed that new oocytes might be originated from ovarian cortical mesenchymal cells. Nayernia *et al*[46–50] assessed the capacity of BM-MSCs to produce male germ cells and claimed that there is a new aspect of germ cell development for the application of BM-MSCs in reproductive medicine. They also indicated that mouse MSCs have the ability for differentiation toward germline stem cells *in vitro* and this relieved that the differentiated cells stop progress at premeiotic stages after transplantation into the testes of mature infertile mice[51]. In another study, Drusenheimer *et al*[52] also demonstrated that spermatozoa can be derived and differentiated from human BM-MSCs.

4. MSCs derived from umbilical cord

The umbilical cord has been located between fetus and mother during pregnancy and which is contained a mucous connective tissue, known as Wharton's jelly, between the amniotic epithelium and the umbilical vessels[53]. Human umbilical cord is a tissue which consists of at least six distinctive zones including from outside to inside: 1) surface epithelium; 2) sub-amniotic stroma; 3) clefts; 4) intervacular stroma also known as Wharton's jelly; 5) perivascular stroma; and 6) vessels (Figure 1)[24]. MSCs have been collected from several parts of the umbilical cord including umbilical cord blood, umbilical vein sub-endothelium, and the Wharton's jelly[24]. MSCs, which are derived from the human umbilical cord (hUC-MSCs), share many traits with BM-MSCs, for instance, they have low expression capacity for HLA major histocompatibility complex class I, self-renewal ability and the capacity to be differentiated into various cell lineages[24], however, they don't have the capacity for expression of CD31, CD45, HLA major histocompatibility complex class I (Table 1)[54]. They also can be frozen/thawed and extensively expanded in culture[39]. Carlin *et al*[55] was the first one who reported the expression of Oct-4, Sox-2, and Nanog markers (some of the embryonic stem cell markers) in porcine umbilical cord matrix cells. A large body of studies indicated that derived hUC-MSCs from extra-embryonic mesoderm, have differentiation potential toward osteogenic, adipogenic, chondrogenic lineages[24,56]. The hUC-MSCs are able to sustain the normal ovarian physiology and decrease the rate of apoptosis in mice model of premature ovarian failure[57].

5. MSCs derived from Wharton's jelly

The primary role of Wharton's jelly is the suppression of compression and torsion and then support of bidirectional blood flow between fetal and maternal circulation and also help the function of adventitia[23,58]. Wharton's jelly is a potential source to be applied in

clinical applications due to their lower risk of viral contamination. MSCs have been isolated from various zones of Wharton's jelly, sub-amnion region, perivascular zone and the intervacular zone (Figure 1). Wharton's jelly-MSCs (WJ-MSCs) have the ability to differentiate toward all three cell lineages. They have the expression profile as same as other MSCs (CD29, CD44, CD73, CD105, CD73, and CD90) and as embryonic stem cell markers such as SSEA-1 and 4, Oct-4, Nanog and Sox-2 (Table 1)[59].

The higher telomerase activity, the higher proliferative potential, shorter expansion doubling times with maintenance of stem cell properties that present in WJ-MSCs in compared with MSCs derived from adult tissues, indicate that WJ-MSCs are in more primitive stage and using them in regenerative medicine has higher privileges[55]. The ability of WJ-MSCs to differentiate toward particular cell lineage depends partially on secreted growth and differentiation factors that are secreted in an environment of a particular cell lineage. Bone morphogenetic protein 4 (BMP4) and retinoic acid (RA) are two other vital factors which their role in differentiation induction has been proved[60]. *In vitro* studies showed that BMP4 induces differentiation of BM-MSCs into primordial germ cells[60]. Moreover, Ohta *et al*[61] have claimed that fetal male germ cells have the machinery to respond RA signals and be differentiated into germ line cells. Applying co-culture system is another safe approach for inducing differentiation of stem cells into specific cell lineage and using them for clinical trial purposes[62].

Deferent studies revealed that WJ-MSCs have the innate capacity, due to an enhanced proliferation potential and a higher rate of colony formation, to be used as an allogeneic cell therapy for diseases treatment[63]. Tamura *et al*[64] indicated that these cells produce several secretory proteins which increase the cancer cells death and stop the cell cycle as well as are markable decrease in the liver fibrosis. There is some evidence that confirms the supportive function of WJ-MSCs for other stem cells. For instance, WJ-MSCs support embryonic germ cell migration by secretion of glial-derived neurotrophic factor, an essential factor to keep the undifferentiated status of spermatogonial stem cells[65].

Asgari *et al*[66] indicated that human WJ-MSCs have the gene expression profile as same as primitive genes in oocyte development after co-culture with placental cells. They reported that supplemented placental cell with transforming growth factor- α and β and basic fibroblast growth factor in a co-culture system is an optimal condition which stimulates hUMSCs to be differentiated toward primordial germ cells and expresses oocyte-like genes. Amidi *et al*[67] reported that in a co-culture system between WJ-MSCs and placenta cells, differentiation potential of MSCs toward male germ-like cell improved when RA and BMP4 were present.

6. MSCs derived from adipose tissue

Just like bone marrow, adipose tissue has been originated from

the mesenchyme and this great source of MSCs has a stroma which can be easily isolated[85]. Adipose tissue is another useful source of multipotent MSCs which called adipose tissue-derived stromal cells (ADSCs). In order to isolate ADSCs, after vigorous digestion and following multiple centrifugation steps, the stromal vascular are isolated[86]. Zuk *et al*[87] reported that ADSCs are similar to BM-MSCs in both differentiation capacities and gene expression[87]. They also reported that ADSCs expression levels of CD49d, CD34, and CD54 are high; however, the expression of CD106 is much higher in BM-MSCs.

7. Conclusion

Wharton's jelly, umbilical cord, and bone marrow are rich sources of MSCs for investigations and presumptive clinical usages. MSCs are ethically reliable, and have a high rate of proliferation and sufficient plasticity for such clinical applications. New progress in cryopreservation methods will open up recent great achievements in MSCs banking and further possibilities for application of cells in regenerative medicine. Meanwhile, Wharton's jelly can be applied in regenerative medicine of some reproductive diseases. These cells have a considerable potency to be differentiated toward germ-like cell lines in appropriate culture condition using BMP4 and RA. Therefore, clinical application of Wharton's jelly has been kept in mind as a promising source for regenerative medicine.

Conflict of interest statement

There is no conflict of interest in the current study.

References

- [1] Caplan AI. Adult mesenchymal stem cells for tissue engineering versus regenerative medicine. *J Cell Physiol* 2007; **213**(2): 341-347.
- [2] Asgari HR, Akbari M, Yazdekhasti H, Rajabi Z, Navid S, Aliakbari F, et al. Comparison of human amniotic, chorionic, and umbilical cord multipotent mesenchymal stem cells regarding their capacity for differentiation toward female germ cells. *Cell Reprogram* 2017; **19**(1): 44-53.
- [3] Dimarino AM, Caplan AI, Bonfield TL. Mesenchymal stem cells in tissue repair. *Front Immunol* 2013; **4**(201): 3389.
- [4] Bonfield TL, Nolan MT, Lennon DP, Caplan AI. Defining human mesenchymal stem cell efficacy *in vivo*. *J Inflamm* 2010; **7**(1): 1.
- [5] Quevedo HC, Hatzistergos KE, Oskouei BN, Feigenbaum GS, Rodriguez JE, Valdes D, et al. Allogeneic mesenchymal stem cells restore cardiac function in chronic ischemic cardiomyopathy via trilineage differentiating capacity. *Proc Natl Acad Sci USA* 2009; **106**(33): 14022-14027.
- [6] Di Nicola M, Carlo-Stella C, Magni M, Milanese M, Longoni PD,

- Matteucci P, et al. Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli. *Blood* 2002; **99**(10): 3838-3843.
- [7] Nasef A, Ashammakhi N, Fouillard L. Immunomodulatory effect of mesenchymal stromal cells: Possible mechanisms. *Regen Med* 2008; **3**(4): 531-546.
- [8] Le Blanc K, Ringden O. Immunomodulation by mesenchymal stem cells and clinical experience. *J Int Med* 2007; **262**(5): 509-525.
- [9] Tian Y, Deng Y, Huang Y, Wang Y. Bone marrow-derived mesenchymal stem cells decrease acute graft-versus-host disease after allogeneic hematopoietic stem cells transplantation. *Immunol Invest* 2007; **37**(1): 29-42.
- [10] Patel SA, Sherman L, Munoz J, Rameshwar P. Immunological properties of mesenchymal stem cells and clinical implications. *Arch Immunol Ther Exp* 2008; **56**(1): 1-8.
- [11] Anzalone R, Iacono ML, Loria T, Di Stefano A, Giannuzzi P, Farina F, et al. Wharton's jelly mesenchymal stem cells as candidates for beta cells regeneration: Extending the differentiative and immunomodulatory benefits of adult mesenchymal stem cells for the treatment of type 1 diabetes. *Stem Cell Rev Rep* 2011; **7**(2): 342-363.
- [12] Abomaray F, Al Jumah M, Kalionis B, Al Askar A, Al Harthy S, Jawdat D, et al. Human chorionic villous mesenchymal stem cells modify the functions of human dendritic cells, and induce an anti-inflammatory phenotype in CD1+ dendritic cells. *Stem Cell Rev Rep* 2015; **11**(3): 423-441.
- [13] Arthur A, Zannettino A, Gronthos S. The therapeutic applications of multipotential mesenchymal/stromal stem cells in skeletal tissue repair. *J Cell Physiol* 2009; **218**(2): 237-245.
- [14] Chen MY, Lie PC, Li ZL, Wei X. Endothelial differentiation of Wharton's jelly-derived mesenchymal stem cells in comparison with bone marrow-derived mesenchymal stem cells. *Exp Hematol* 2009; **37**(5): 629-640.
- [15] Yazdekhasti H, Rajabi Z, Parvari S, Abbasi M. Used protocols for isolation and propagation of ovarian stem cells, different cells with different traits. *J Ovarian Res* 2016; **9**(1): 68.
- [16] Yazdekhasti H, Rajabi Z, Akrami SM. Ethical issues associated with advanced paternal age and genetic disorders in their offspring. *Arvand J Health Med Sci* 2016; **1**(4): 191-198.
- [17] Rezaeian Z, Yazdekhasti H, Nasri S, Rajabi Z, Fallahi P, Amidi F. Effect of selenium on human sperm parameters after freezing and thawing procedures. *Asian Pac J Reprod* 2016; **5**(6): 462-466.
- [18] Najar M, Raicevic G, Boufker HI, Kazan HF, De Bruyn C, Meuleman N, et al. Mesenchymal stromal cells use PGE2 to modulate activation and proliferation of lymphocyte subsets: Combined comparison of adipose tissue, Wharton's Jelly and bone marrow sources. *Cell Immunol* 2010; **264**(2): 171-179.
- [19] Bobis S, Jarocho D, Majka M. Mesenchymal stem cells: Characteristics and clinical applications. *Folia Histochem Cytobiol* 2006; **44**(4): 215.
- [20] Guillot PV, Gotherstrom C, Chan J, Kurata H, Fisk NM. Human first-trimester fetal MSC express pluripotency markers and grow faster and have longer telomeres than adult MSC. *Stem Cells* 2007; **25**(3): 646-654.
- [21] Götherström C, Ringden O, Westgren M, Tammik C, Le Blanc K. Immunomodulatory effects of human foetal liver-derived mesenchymal stem cells. *Bone Marrow Transplant* 2004; **33**(11): 1167-1167.
- [22] Troyer DL, Weiss ML. Concise review: Wharton's Jelly-derived cells are a primitive stromal cell population. *Stem Cells* 2008; **26**(3): 591-599.
- [23] Can A, Karahuseyinoglu S. Concise review: Human umbilical cord stroma with regard to the source of fetus-derived stem cells. *Stem Cells* 2007; **25**(11): 2886-2895.
- [24] Karahuseyinoglu S, Cinar O, Kilic E, Kara F, Akay GG, Demiralp DÖ, et al. Biology of stem cells in human umbilical cord stroma: in situ and in vitro surveys. *Stem Cells* 2007; **25**(2): 319-331.
- [25] Bailey MM, Wang L, Bode CJ, Mitchell KE, Detamore MS. A comparison of human umbilical cord matrix stem cells and temporomandibular joint condylar chondrocytes for tissue engineering temporomandibular joint condylar cartilage. *Tissue Eng* 2007; **13**(8): 2003-2010.
- [26] Jomura S, Uy M, Mitchell K, Dallasen R, Bode CJ, Xu Y. Potential treatment of cerebral global ischemia with oct-4+ umbilical cord matrix cells. *Stem Cells* 2007; **25**(1): 98-106.
- [27] Fu YS, Cheng YC, Lin MYA, Cheng H, Chu PM, Chou SC, et al. Conversion of human umbilical cord mesenchymal stem cells in Wharton's jelly to dopaminergic neurons in vitro: Potential therapeutic application for Parkinsonism. *Stem Cells* 2006; **24**(1): 115-124.
- [28] Mitchell KE, Weiss ML, Mitchell BM, Martin P, Davis D, Morales L, et al. Matrix cells from Wharton's jelly form neurons and glia. *Stem Cells* 2003; **21**(1): 50-60.
- [29] Romanov YA, Svintsitskaya VA, Smirnov VN. Searching for alternative sources of postnatal human mesenchymal stem cells: Candidate MSC-like cells from umbilical cord. *Stem Cells* 2003; **21**(1): 105-110.
- [30] Atala A. *Methods of tissue engineering*. Houston: Gulf Professional Publishing; 2002.
- [31] Seward L, Zahradka P. Coronary artery smooth muscle in culture: Migration of heterogeneous cell populations from vessel wall. *Mol Cell Biochem* 1997; **176**(1-2): 53-59.
- [32] Isakson M, de Blacam C, Whelan D, McArdle A, Clover A. Mesenchymal stem cells and cutaneous wound healing: Current evidence and future potential. *Stem Cells Int* 2015; **2015**: 831095.
- [33] Friedenstien AJ, Gorskaja J, Kulagina N. Fibroblast precursors in normal and irradiated mouse hematopoietic organs. *Exp Hematol* 1976; **4**(5): 267-274.
- [34] Caplan AI. Mesenchymal stem cells. *J Orthopaed Res* 1991; **9**(5): 641-650.
- [35] Horwitz E, Le Blanc K, Dominici M, Mueller I, Slaper-Cortenbach I, Marini FC, et al. Clarification of the nomenclature for MSC: The international society for cellular therapy position statement. *Cytotherapy* 2005; **7**(5): 393-395.
- [36] Logeart-Avramoglou D, Anagnostou F, Bizios R, Petite H. Engineering bone: Challenges and obstacles. *J Cellul Mol Med* 2005; **9**(1): 72-84.
- [37] Kadivar M, Khatami S, Mortazavi Y, Soleimani M, Taghikhani M, Shokrgozar MA. Isolation, culture and characterization of postnatal human umbilical vein-derived mesenchymal stem cells. *DARU J Pharmaceut Sci* 2005; **13**(4): 170-176.

- [38]Secco M, Moreira YB, Zucconi E, Vieira NM, Jazedje T, Muotri AR, et al. Gene expression profile of mesenchymal stem cells from paired umbilical cord units: Cord is different from blood. *Stem Cell Rev Rep* 2009; **5**(4): 387-401.
- [39]Anzalone R, Iacono ML, Corrao S, Magno F, Loria T, Cappello F, et al. New emerging potentials for human Wharton's jelly mesenchymal stem cells: Immunological features and hepatocyte-like differentiative capacity. *Stem Cells Dev* 2010; **19**(4): 423-438.
- [40]Tipnis S, Viswanathan C, Majumdar AS. Immunosuppressive properties of human umbilical cord-derived mesenchymal stem cells: Role of B7-H1 and IDO. *Immunol Cell Biol* 2010; **88**(8): 795-806.
- [41]Corcione A, Benvenuto F, Ferretti E, Giunti D, Cappiello V, Cazzanti F, et al. Human mesenchymal stem cells modulate B-cell functions. *Blood* 2006; **107**(1): 367-372.
- [42]Sotiropoulou PA, Perez SA, Gritzapis AD, Baxeavanis CN, Papamichail M. Interactions between human mesenchymal stem cells and natural killer cells. *Stem Cells* 2006; **24**(1): 74-85.
- [43]Johnson J, Bagley J, Skaznik-Wikiel M, Lee HJ, Adams GB, Niikura Y, et al. Oocyte generation in adult mammalian ovaries by putative germ cells in bone marrow and peripheral blood. *Cell* 2005; **122**(2): 303-315.
- [44]Bukovsky A, Svetlikova M, Caudle MR. Oogenesis in cultures derived from adult human ovaries. *Reprod Biol Endocrinol* 2005; **3**(1): 1.
- [45]Yazdekhasti H, Hosseini MA, Rajabi Z, Parvari S, Salehnia M, Koruji M, et al. Improved isolation, proliferation, and differentiation capacity of mouse ovarian putative stem cells. *Cell Rerogram* 2017; **19**(2): 132-144.
- [46]Nayernia K, Lee JH, Drusenheimer N, Nolte J, Wulf G, Dressel R, et al. Derivation of male germ cells from bone marrow stem cells. *Lab Invest* 2006; **86**(7): 654-663.
- [47]Aliakbari F, Gilani MAS, Amidi F, Baazm M, Korouji M, Izadyar F, et al. Improving the efficacy of cryopreservation of spermatogonia stem cells by antioxidant supplements. *Cell Rerogram* 2016; **18**(2): 87-95.
- [48]Aliakbari F, Yazdekhasti H, Abbasi M, Hajian Monfared M, Baazm M. Advances in cryopreservation of spermatogonial stem cells and restoration of male fertility. *Microsc Res Tech* 2016; **79**(2): 122-129.
- [49]Parvari S, Yazdekhasti H, Rajabi Z, Gerayeli Malek V, Rastegar T, Abbasi M. Differentiation of mouse ovarian stem cells toward oocyte-like structure by coculture with granulosa cells. *Cell Rerogram* 2016; **18**(6): 419-428.
- [50]Aliakbari F, Sedighi Gilani MA, Yazdekhasti H, Koruji M, Asgari HR, Baazm M, et al. Effects of antioxidants, catalase and α -tocopherol on cell viability and oxidative stress variables in frozen-thawed mice spermatogonial stem cells. *Artif Cells Nanomed Biotechnol* 2017; **45**(1): 63-68.
- [51]Nayernia K, Li M, Jaroszynski L, Khusainov R, Wulf G, Schwandt I, et al. Stem cell based therapeutical approach of male infertility by teratocarcinoma derived germ cells. *Hum Mol Genet* 2004; **13**(14): 1451-1460.
- [52]Drusenheimer N, Wulf G, Nolte J, Lee JH, Dev A, Dressel R, et al. Putative human male germ cells from bone marrow stem cells. *Soc Reprod Fertil Suppl* 2007; **63**: 69.
- [53]Baksh D, Yao R, Tuan RS. Comparison of proliferative and multilineage differentiation potential of human mesenchymal stem cells derived from umbilical cord and bone marrow. *Stem Cells* 2007; **25**(6): 1384-1392.
- [54]Weiss ML, Anderson C, Medicetty S, Seshareddy KB, Weiss RJ, VanderWerff I, et al. Immune properties of human umbilical cord Wharton's jelly-derived cells. *Stem Cells* 2008; **26**(11): 2865-2874.
- [55]Carlin R, Davis D, Weiss M, Schultz B, Troyer D. Expression of early transcription factors Oct-4, Sox-2 and Nanog by porcine umbilical cord (PUC) matrix cells. *Reprod Biol Endocrinol* 2006; **4**(1): 8.
- [56]Karahuseyinoglu S, Kocaefe C, Balci D, Erdemli E, Can A. Functional structure of adipocytes differentiated from human umbilical cord stroma-derived stem cells. *Stem Cells* 2008; **26**(3): 682-691.
- [57]Miyamoto K, Hayashi K, Suzuki T, Ichihara S, Yamada T, Kano Y, et al. Human placenta feeder layers support undifferentiated growth of primate embryonic stem cells. *Stem Cells* 2004; **22**(4): 433-440.
- [58]Petsa A, Gargani S, Felesakis A, Grigoriadis N, Grigoriadis I. Effectiveness of protocol for the isolation of Wharton's Jelly stem cells in large-scale applications. *In Vitro Cell Dev Biol Anim* 2009; **45**(10): 573-576.
- [59]Taghizadeh R, Cetrulo K, Cetrulo C. Wharton's Jelly stem cells: Future clinical applications. *Placenta* 2011; **32**: S311-S315.
- [60]Ewen-Campen B, Schwager EE, Extavour CG. The molecular machinery of germ line specification. *Mol Reprod Dev* 2010; **77**(1): 3-18.
- [61]Ohta K, Lin Y, Hogg N, Yamamoto M, Yamazaki Y. Direct effects of retinoic acid on entry of fetal male germ cells into meiosis in mice. *Biol Reprod* 2010; **83**(6): 1056-1063.
- [62]Iwahashi K, Yoshioka H, Low EW, McCarrey JR, Yanagimachi R, Yamazaki Y. Autonomous regulation of sex-specific developmental programming in mouse fetal germ cells. *Biol Reprod* 2007; **77**(4): 697-706.
- [63]Hendijani F, Sadeghi-Aliabadi H, Javanmard SH. Comparison of human mesenchymal stem cells isolated by explant culture method from entire umbilical cord and Wharton's jelly matrix. *Cell Tissue Bank* 2014; **15**(4): 555-565.
- [64]Watson N, Divers R, Kedar R, Mehindru A, Mehindru A, Borlongan MC, et al. Discarded Wharton jelly of the human umbilical cord: A viable source for mesenchymal stromal cells. *Cytotherapy* 2015; **17**(1): 18-24.
- [65]Kubota H, Avarbock MR, Brinster RL. Growth factors essential for self-renewal and expansion of mouse spermatogonial stem cells. *Proc Natl Acad Sci USA* 2004; **101**(47): 16489-16494.
- [66]Asgari HR, Akbari M, Abbasi M, Ai J, Korouji M, Aliakbari F, et al. Human Wharton's jelly-derived mesenchymal stem cells express oocyte developmental genes during co-culture with placental cells. *Iran J Basic Med Sci* 2015; **18**(1): 22.
- [67]Amidi F, Hoseini MA, Nia KN, Habibi M, Kajbafzadeh AM, Mazaheri Z, et al. Male germ-like cell differentiation potential of human umbilical cord Wharton's jelly-derived mesenchymal stem cells in co-culture with human placenta cells in presence of BMP4 and retinoic acid. *Iran J Basic Med Sci* 2015; **18**(4): 325-333.
- [68]Turnovcova K, Ruzickova K, Vanecek V, Sykova E, Jendelova P. Properties and growth of human bone marrow mesenchymal stromal cells cultivated in different media. *Cytotherapy* 2009; **11**(7): 874-885.

- [69]La Rocca G, Anzalone R, Corrao S, Magno F, Loria T, Iacono ML, et al. Isolation and characterization of Oct-4+/HLA-G+ mesenchymal stem cells from human umbilical cord matrix: Differentiation potential and detection of new markers. *Histochem Cell Biol* 2009; **131**(2): 267-282.
- [70]Weiss ML, Medicetty S, Bledsoe AR, Rachakatla RS, Choi M, Merchav S, et al. Human umbilical cord matrix stem cells: Preliminary characterization and effect of transplantation in a rodent model of Parkinson's disease. *Stem Cells* 2006; **24**(3): 781-792.
- [71]Wang HS, Hung SC, Peng ST, Huang CC, Wei HM, Guo YJ, et al. Mesenchymal stem cells in the Wharton's jelly of the human umbilical cord. *Stem Cells* 2004; **22**(7): 1330-1337.
- [72]Lu LL, Liu YJ, Yang SG, Zhao QJ, Wang X, Gong W, et al. Isolation and characterization of human umbilical cord mesenchymal stem cells with hematopoiesis-supportive function and other potentials. *Haematologica* 2006; **91**(8): 1017-1026.
- [73]Lund RD, Wang S, Lu B, Girman S, Holmes T, Sauve Y, et al. Cells isolated from umbilical cord tissue rescue photoreceptors and visual functions in a rodent model of retinal disease. *Stem Cells* 2007; **25**(3): 602-611.
- [74]Wu KH, Zhou B, Lu SH, Feng B, Yang SG, Du WT, et al. *In vitro* and *in vivo* differentiation of human umbilical cord derived stem cells into endothelial cells. *J Cell Biochem* 2007; **100**(3): 608-616.
- [75]Lupatov AY, Karalkin P, Suzdal'tseva YG, Burunova V, Yarygin V, Yarygin K. Cytofluorometric analysis of phenotypes of human bone marrow and umbilical fibroblast-like cells. *Bull Exp Biol Med* 2006; **142**(4): 521-526.
- [76]Kadner A, Zund G, Maurus C, Breymann C, Yakarisik S, Kadner G, et al. Human umbilical cord cells for cardiovascular tissue engineering: A comparative study. *Eur J Cardiothorac Surg* 2004; **25**(4): 635-641.
- [77]Selmani Z, Naji A, Zidi I, Favier B, Gaiffe E, Obert L, et al. Human leukocyte antigen-G5 secretion by human mesenchymal stem cells is required to suppress T lymphocyte and natural killer function and to induce CD4+ CD25 high FOXP3+ regulatory T cells. *Stem Cells* 2008; **26**(1): 212-222.
- [78]Sarugasar R, Lickorish D, Baksh D, Hosseini MM, Davies JE. Human umbilical cord perivascular (HUCPV) cells: A source of mesenchymal progenitors. *Stem Cells* 2005; **23**(2): 220-229.
- [79]Kadivar M, Khatami S, Mortazavi Y, Shokrgozar MA, Taghikhani M, Soleimani M. *In vitro* cardiomyogenic potential of human umbilical vein-derived mesenchymal stem cells. *Biochem Biophys Res Commun* 2006; **340**(2): 639-647.
- [80]Ren G, Zhao X, Zhang L, Zhang J, L'Huillier A, Ling W, et al. Inflammatory cytokine-induced intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 in mesenchymal stem cells are critical for immunosuppression. *J Immunol* 2010; **184**(5): 2321-2328.
- [81]Gotherstrom C, West A, Liden J, Uzunel M, Lahesmaa R, Le Blanc K. Difference in gene expression between human fetal liver and adult bone marrow mesenchymal stem cells. *Haematologica* 2005; **90**(8): 1017-1026.
- [82]Guo Z, Zheng C, Chen Z, Gu D, Du W, Ge J, et al. Fetal BM-derived mesenchymal stem cells promote the expansion of human Th17 cells, but inhibit the production of Th1 cells. *Eur J Immunol* 2009; **39**(10): 2840-2849.
- [83]Deans RJ, Moseley AB. Mesenchymal stem cells: Biology and potential clinical uses. *Exp Hematol* 2000; **28**(8): 875-884.
- [84]Carlin R, Davis D, Weiss M, Schultz B, Troyer D. Expression of early transcription factors Oct-4, Sox-2 and Nanog by porcine umbilical cord (PUC) matrix cells. *Reprod Biol Endocrinol* 2006; **4**(1): 1.
- [85]Wei G, Schubiger G, Harder F, Müller AM. Stem cell plasticity in mammals and transdetermination in Drosophila: Common themes? *Stem Cells* 2000; **18**(6): 409-414.
- [86]McArdle A, Chung MT, Paik KJ, Duldulao C, Chan C, Rennert R, et al. Positive selection for bone morphogenetic protein receptor type-IB promotes differentiation and specification of human adipose-derived stromal cells toward an osteogenic lineage. *Tissue Eng Part A* 2014; **20**(21-22): 3031-3040.
- [87]Zuk PA, Zhu M, Ashjian P, De Ugarte DA, Huang JI, Mizuno H, et al. Human adipose tissue is a source of multipotent stem cells. *Mol Biol Cell* 2002; **13**(12): 4279-4295.